

# Infections in Transplant Recipients

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Infectious Disease

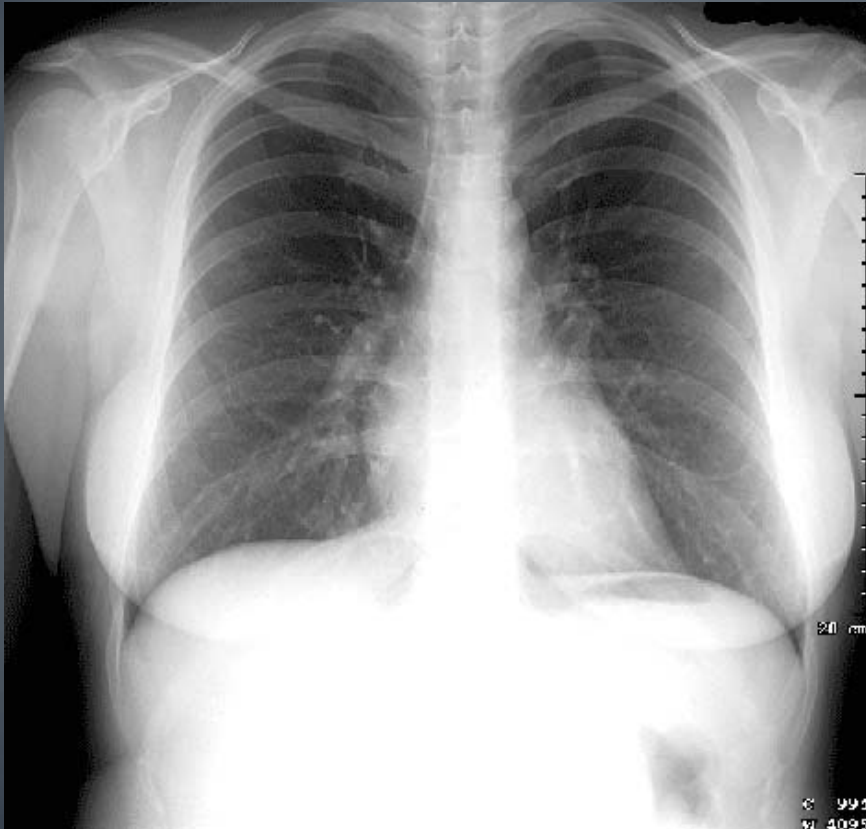
# Objectives

- Identify important aspects of the pre-transplant infectious workup
- Identify infections associated with transplant recipients
- Recognize the role and timing immunosuppression plays in transplant related infections

# Case #1

- A 47 year old female presents to your office for evaluation prior to renal transplant for diabetic nephropathy. PMHx is significant for HTN, DM. She was born in Mexico and recalls her Aunt having tuberculosis when she was a child. She has no cough, fever, chills, night sweats, weight loss or other complaints. CXR is ordered. Quantiferon is ordered and the results are below:
  - Mitogen >10.00
  - Nil 0.02
  - Tb Ag 1 – Nil 7.51
  - TB Ag 2 – Nil 6.13
- How do you interpret this result?

# Case #1



Her CXR is back. Based on this information, what is your next step?

- A) Send sputum for AFB x3
- B) Clear her for transplant at this time
- C) Start RIF 600mg daily for 4 months
- D) Start RIPE daily for 2 months, following by 4 months of INH/RIF/B6 and hold transplant until complete?

# Pretransplant

- Pretransplant history of utmost importance
  - Prior exposures, travel, occupations, hobbies
- Chronic diseases may be affected by transplant
  - ?re-infection of transplanted organ (HBV, HCV)
  - Diabetes mellitus affecting graft healing
  - Pre-existing cardiac disease, pulmonary, gall bladder
- Prior exposures/immunity play a role in immunosuppressed persons
- Prior colonization/infections
- Reactivation of latent infection

**TABLE 311-5** Routine Laboratory Studies before and after Transplantation

| BEFORE TRANSPLANTATION*   | AFTER TRANSPLANTATION                      |
|---|--|
| Cytomegalovirus IgG antibody  | Viral load monitoring for cytomegalovirus  |
| Epstein-Barr virus IgG antibody   | Antibody studies (as clinically indicated) |
| Herpes simplex (types 1 and 2) antibody                                 |  |
| Varicella-zoster IgG antibody   |  |
| <i>Toxoplasma</i> IgG antibody (heart transplant recipients)            |  |
| Hepatitis B screen <sup>†</sup>   |  |
| Hepatitis C enzyme immunoassay <sup>‡</sup>                             |  |
| Human immunodeficiency virus antibody                                   |  |
| Tuberculin skin test or interferon gamma release assay for tuberculosis |  |
| Stool for ova and parasites; <i>Strongyloides</i> antibody <sup>§</sup> |  |
| <i>Trypanosoma cruzi</i> antibody <sup>  </sup>                         |  |

Mandell: Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 8<sup>th</sup> ed.; Chapter 311: Risk Factors and Approaches to Infections in Transplant Recipients. 3414-3424.

# Risk factors for infection

- Patients are undergoing major surgery, many will be in ICU setting post operatively
- Organ was outside the body for hours
  - Ischemia leading to allograft injury?
- Many patients have been in the ICU setting prior to transplantation
- ?Immune suppression making the patient more susceptible to infection (BMT)
- SOT infections are most common at the site of transplantation
- Surgical procedure itself
  - OR time, blood loss, etc.

# CASE #2

A 57 year old female undergoes orthotopic liver transplantation for HCV related cirrhosis. Due to her medical condition, she has been an inpatient for over one month and was in the ICU for several weeks prior to her transplant. 13 days after her transplant, she becomes febrile to 101.1F and her WBC rises to 18.7K. She has been on an empiric course of vancomycin and zosyn since the transplant. Blood cultures turn positive for Gram + cocci in pairs and chains. What is your next course of action?

- A) Repeat blood cultures and remove/exchange any central lines. Await ID of the isolate.
- B) Repeat blood cultures and order a CT of the abd/pelvis. Change her antibiotics to include VRE activity.
- C) Repeat blood cultures, continue vancomycin and zosyn. Add linezolid.
- D) Review donor cultures and information. Repeat blood cultures. Order MRSA swab of nares. Continue your current antibiotics.

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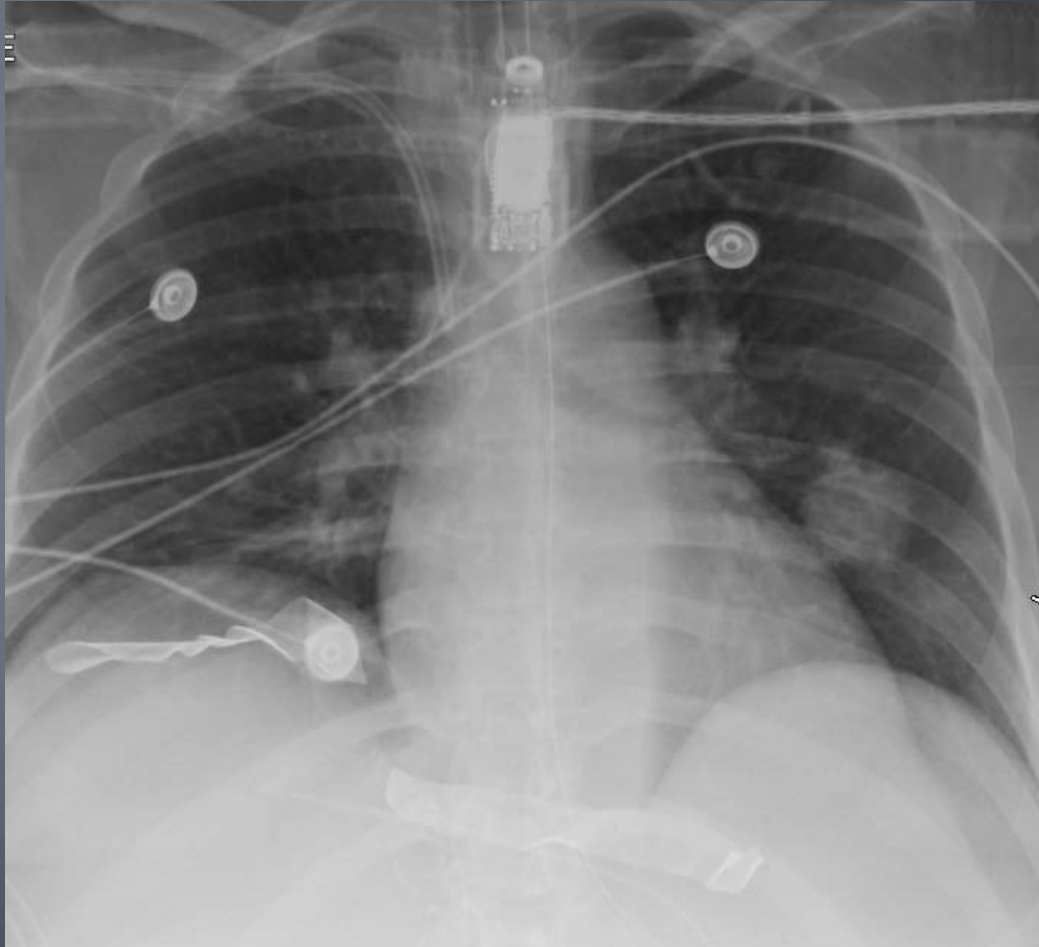
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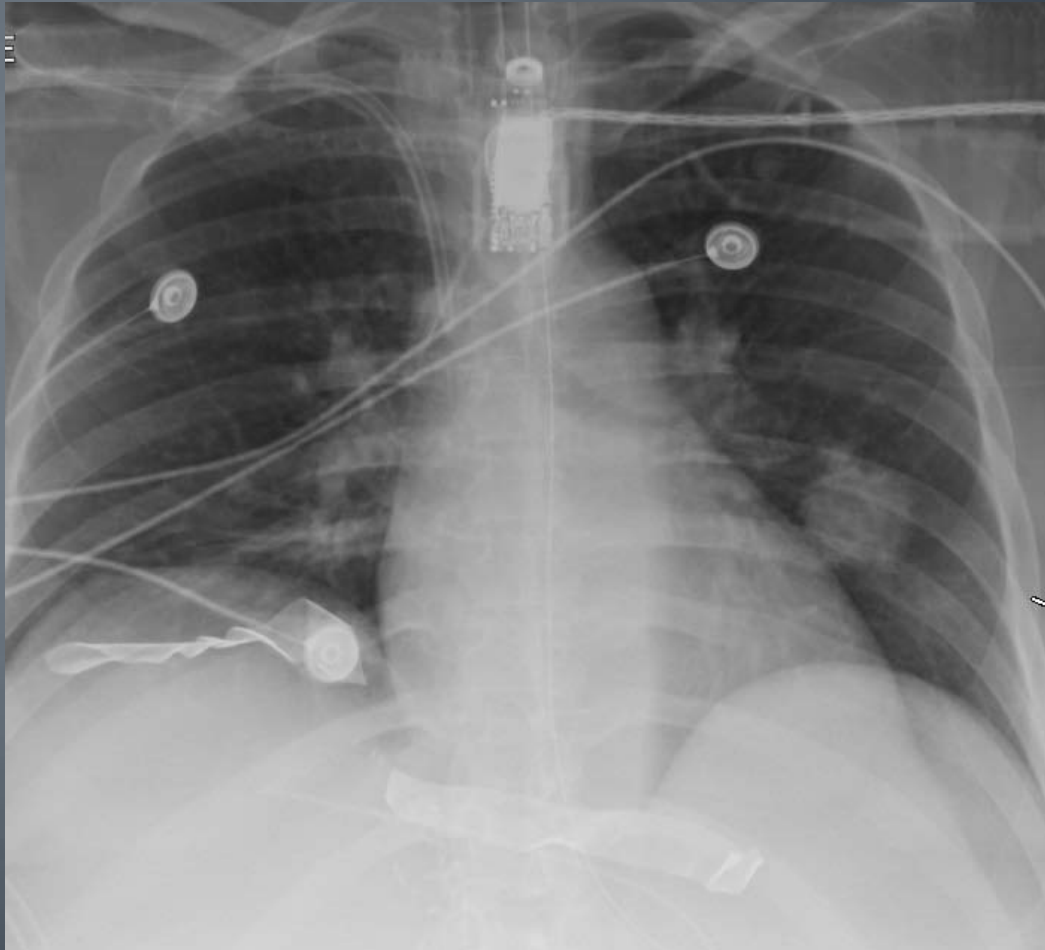
## Case #2

- Your patient recovers from her VRE bacteremia and wound infection. 3 weeks later she is discharged from the hospital. One week following this, she is admitted with one day of LLQ abdominal pain, fever, diarrhea. Temp 101.7F, P109, BP 121/75. She has left lower abdominal pain on exam. WBC is 18.9K.
  - What is the most likely diagnosis?
    - A) Wound infection
    - B) Sepsis of unknown source
    - C) Clostridium difficile colitis
    - D) Medication toxicity

A 57 year old female undergoes orthotopic liver transplantation for HCV related cirrhosis at BUMCP. Due to her medical condition, she has been an inpatient for over one month and was in the ICU for several weeks prior to her transplant. 61 days after her transplant, she becomes febrile to 100.8F. She is currently being treated for VRE bacteremia and candida albicans fungemia with daptomycin and micafungin. She is not on any O2 and does not have a cough but you order a CXR, which is shown below. She takes tacrolimus, MMF, prednisone for IS. She takes Bactrim and valganciclovir for prophylaxis.



You notice she was previously positive for *Coccidioides* IgG and has been off fluconazole for one month now due to starting micafungin.



# What is your suspected diagnosis?

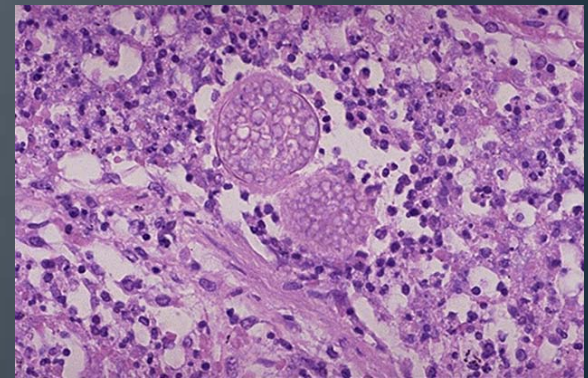
- A) Round atelectasis
- B) Aspergillus pneumonia
- C) Pulmonary Coccidioides
- D) Pulmonary Nocardiosis
- E) MRSA pneumonia

# Coccidioides species

- Dimorphic – existing as either mycelium or spherule
- Two species – *C. immitis* and *C. posadasii*
  - *C. immitis* from California San Joaquin Valley
  - *C. posadasii* – all other endemic areas
- 0.5% cases are disseminated disease
- Pulmonary lesions, skin manifestations
- Prophylaxis for SOT 200mg daily 6-12 months
- Treatment for life after tx course for active disease
- AmB for severe infection



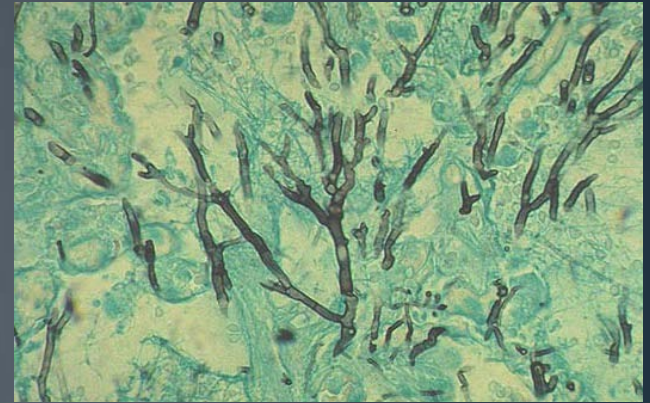
[http://phil.cdc.gov/PHIL/Images/11121998/00004/39G0040\\_lores.jpg](http://phil.cdc.gov/PHIL/Images/11121998/00004/39G0040_lores.jpg)



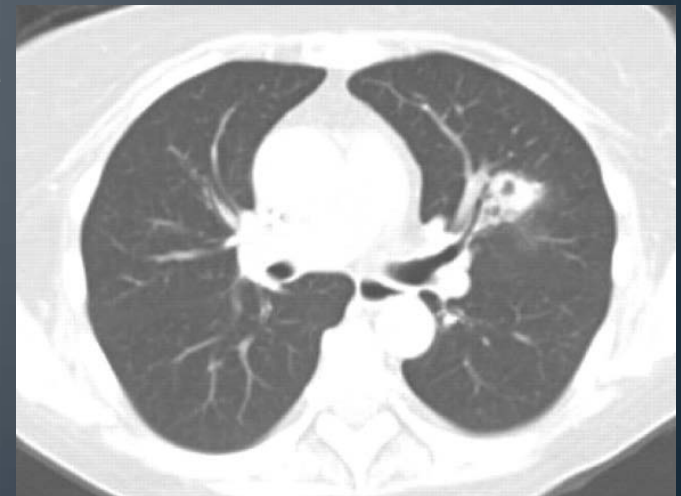
<http://library.med.utah.edu/WebPath/INFEHTML/INFE024.html>

# Aspergillus species

- Major cause morbidity/mortality
- Grow on most media within 48 hours
- Acute angle branching, true septate hyphae
- Aspergillus Ag (Galactomannan) used to detect cell wall for invasive Aspergillosis
  - Not recommended for SOT – only BMT
  - Need tissue/fluid for culture/path
  - Causes wide spectrum of clinical syndromes
- Treat empirically with voriconazole or posaconazole if suspected
- Interactions with IS meds (raises tacrolimus levels)
- Be aware of azole-resistant species

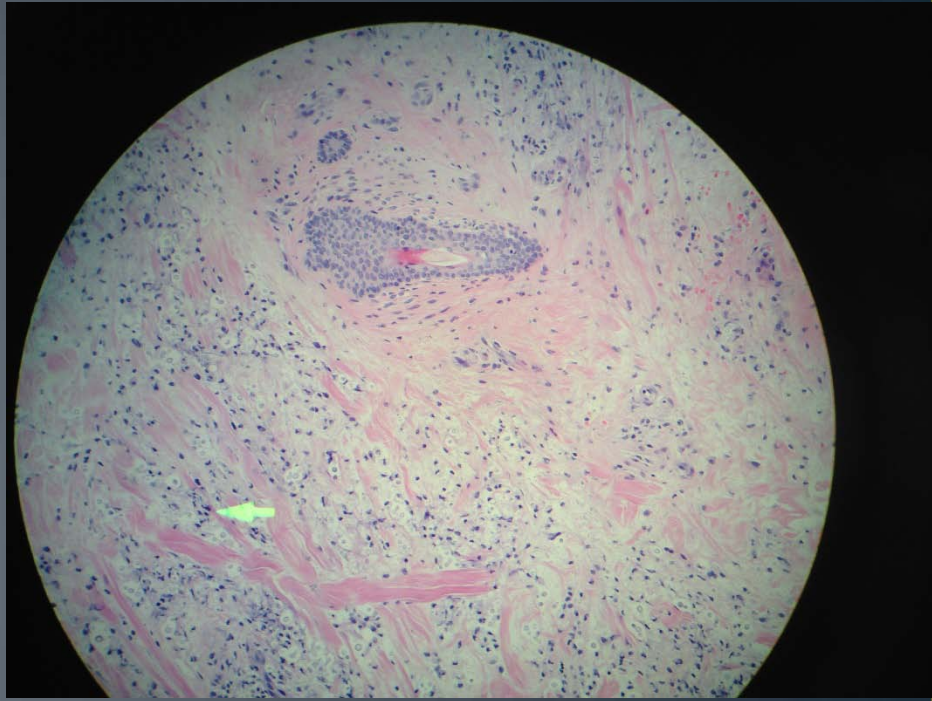


<http://www.pathologyoutlines.com/topic/liveraspergillus.html>



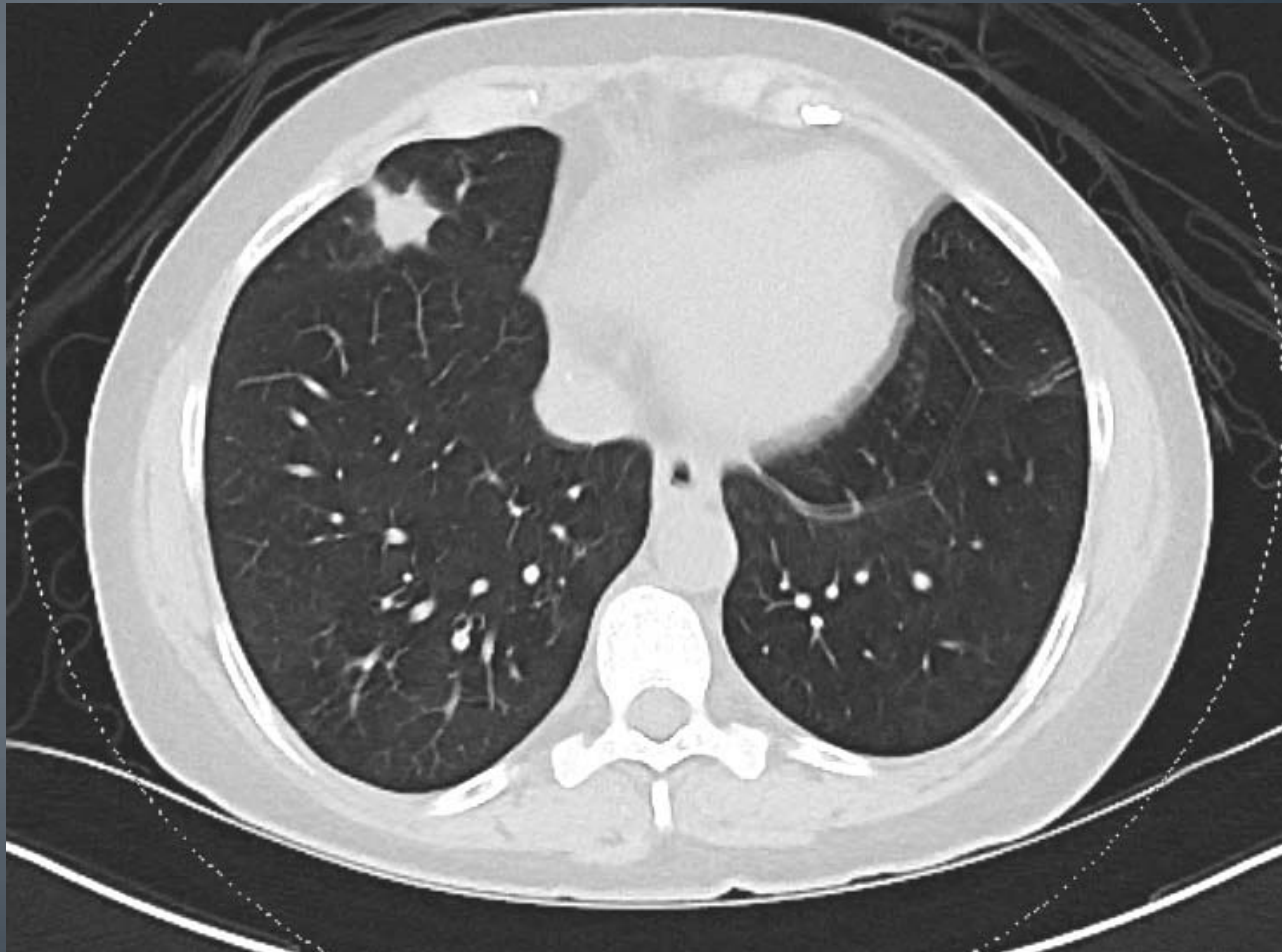
Pulmonary aspergillosis: a clinical review  
M. Kousha, R. Tadi, A.O. Soubani  
European Respiratory Review 2011 20: 156-174;

# Cryptococcus neoformans



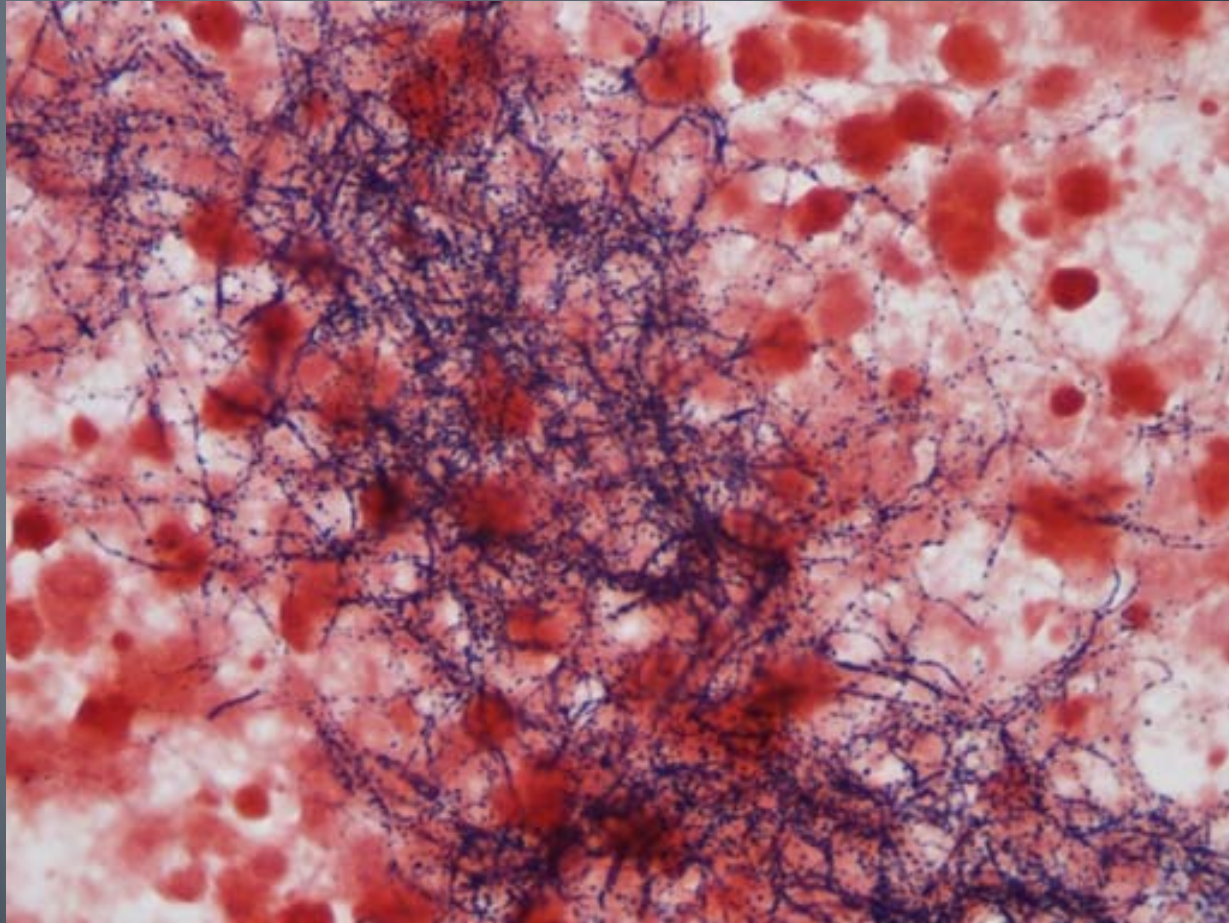
- Common immune compromised pathogen
- Encapsulated
- Associated with pigeons, woods
- Grow on most agar in 2-3 days
- Wide manifestations – most common CNS, pulmonary, cutaneous
- Serum/CSF cryptococcal antigen useful in diagnosis
- Treat disseminated disease like CNS – Amphotericin/flucytosine initially

A 37 year old female is admitted for fatigue, nausea and fever. She previously underwent a renal/pancreas transplant which failed. She underwent a second renal transplant 18 months ago and received campath at the time of her transplant. She has not been treated for rejection. She takes tacrolimus, MMF and valganciclovir. On admission she has a CT chest show below:





While performing your incredibly thorough physical exam, you note a mass in her posterior thigh that is mobile, non tender and without signs of inflammation. An u/s of this reveals concern for abscess. IR aspirates this collection and sends the fluid for gram stain and culture. Gram stain is below:



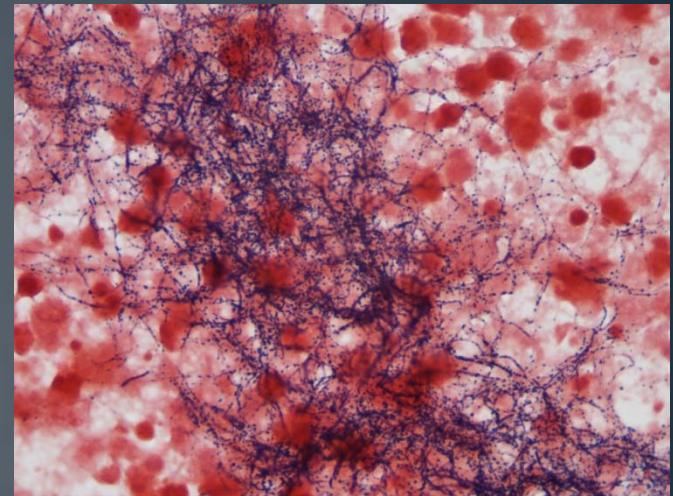
How do you interpret this gram stain?

# What antibiotics do you empirically start?

- A) Vancomycin and Zosyn
- B) Linezolid and Zosyn
- C) Ambisome
- D) Bactrim and Imipenem
- E) Vancomycin, Cefepime and Tobramycin

# *Nocardia* species

- Ubiquitous environmental organism
- Gram positive beaded branching rods
- Direct inoculation or inhalation causes infection
- Many times subacute/chronic presentation
- Pulmonary, cutaneous, CNS disease
- CNS imaging in all cases of pulmonary disease
- Bactrim mainstay of treatment, also carbapenem, amikacin, linezolid
- Prolonged course of treatment
- Less common due to Bactrim prophylaxis



# Case #5

A 32 year old male is hospitalized for a 4 month history of fevers, night sweats and increasing fatigue. Previously he underwent b/l lung transplantation 6 years ago for cystic fibrosis. He is taking MMF, tacrolimus and prednisone. Physical exam is notable for fever 100.7F, P71, BP 127/78. Diffuse lymphadenopathy is noted on exam.

Which of the following tests would assist you in obtaining his diagnosis?

- A) CMV pcr
- B) CMV IgM/IgG
- C) EBV pcr
- D) EBV IgM/IgG
- E) Sputum AFB x3

# EBV (HHV4)

- Primary infection asymptomatic or mononucleosis syndrome (cervical LAD, splenomegaly, pharyngitis)
- 90% have immunity/past infection by age 40
- Post transplant lymphoproliferative disorder (PTLD) spectrum of disease.
  - Polyclonal B cell mononucleosis syndrome to malignant monoclonal lymphoma
- 3-10% of SOT recipients
- Mortality 40-60%
- Suspect with EBV pcr positive
- Confirm diagnosis with tissue biopsy
- Treatment is chemotherapy – usually rituxan. No active antiviral therapy

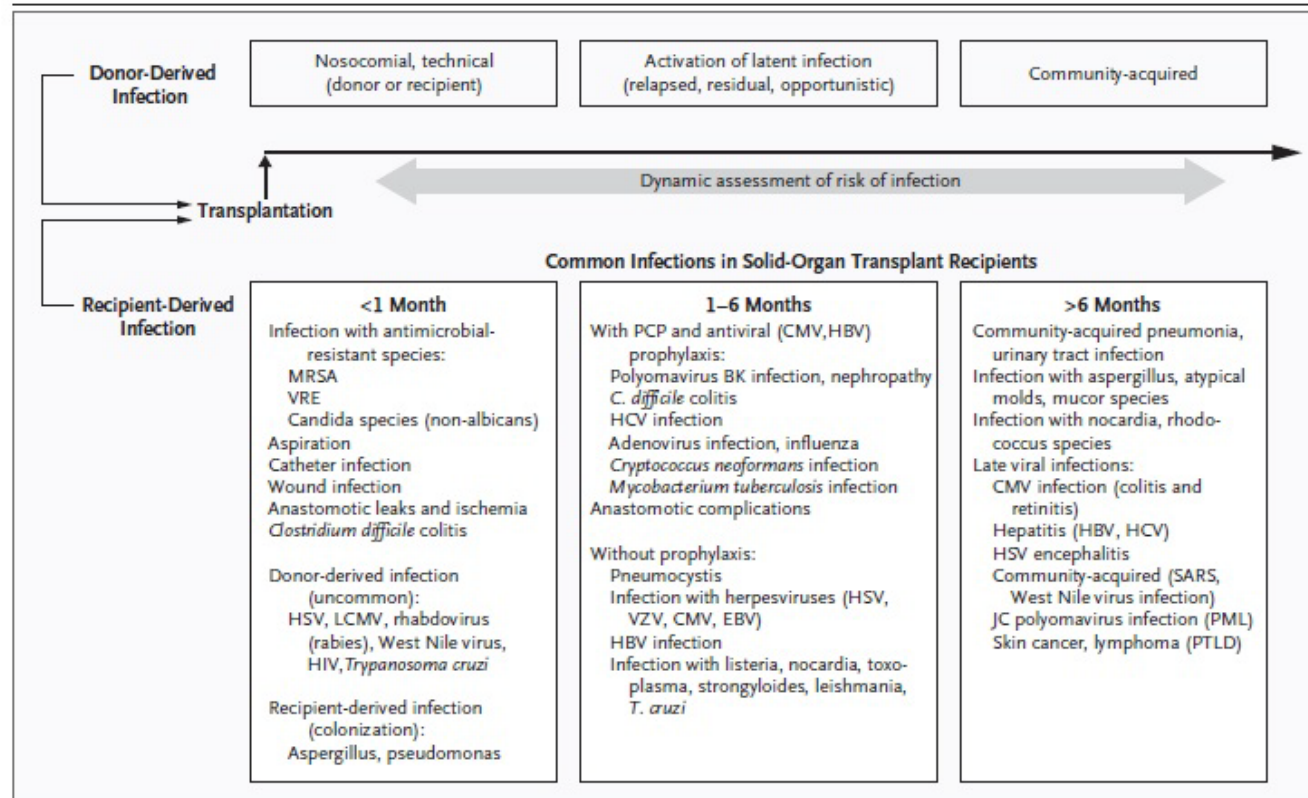
**Table 1. Clinical Presentations of Post-Transplantation Lymphoproliferative Disorder Associated with Epstein-Barr Virus.**

|  |
|--|
| Unexplained fever (fever of unknown origin)                            |
| Mononucleosis-like syndrome (fever, malaise, pharyngitis, tonsillitis) |
| Gastrointestinal bleeding, obstruction, or perforation                 |
| Abdominal-mass lesions   |
| Infiltrative disease of the allograft                                  |
| Hepatocellular or pancreatic dysfunction                               |
| Central nervous system disease   |

Fishman, J. *Infection in Solid-Organ Transplant Recipients.* N Engl J Med 2007;357:2601-14.

# Timeline of Infection

The NEW ENGLAND JOURNAL of MEDICINE



**Figure 4. Changing Timeline of Infection after Organ Transplantation.**

Infections occur in a generally predictable pattern after solid-organ transplantation. The development of infection is delayed by prophylaxis and accelerated by intensified immunosuppression, drug toxic effects that may cause leukopenia, or immunomodulatory viral infections such as infection with cytomegalovirus (CMV), hepatitis C virus (HCV), or Epstein-Barr virus (EBV). At the time of transplantation, a patient's short-term and long-term risk of infection can be stratified according to donor and recipient screening, the technical outcome of surgery, and the intensity of immunosuppression required to prevent graft rejection. Subsequently, an ongoing assessment of the risk of infection is used to adjust both prophylaxis and immunosuppressive therapy. MRSA denotes methicillin-resistant *Staphylococcus aureus*, VRE vancomycin-resistant *Enterococcus faecalis*, HSV herpes simplex virus, LCMV lymphocytic choriomeningitis virus, HIV human immunodeficiency virus, PCP *Pneumocystis carinii* pneumonia, HBV hepatitis B virus, VZV varicella-zoster virus, SARS severe acute respiratory syndrome, PML progressive multifocal leukoencephalopathy, and PTLD post-transplantation lymphoproliferative disorder. Modified from Fishman and Rubin<sup>1</sup> and Rubin et al.<sup>45</sup>

# Recipient derived infections

- Active infections should be eradicated/controlled prior to transplant
  - Immune suppression worsens this
- Endogenous flora
  - Candida
  - VRE
  - ?Staphylococcus aureus/MRSA
  - Aspergillus
  - Pseudomonas
- Latent infections
  - Toxoplasma
  - Herpes viruses
  - Tuberculosis
  - Coccidioides, Histoplasmosis

# Donor derived infections

- Donor information made available to physicians
- Generally uncommon but still do occur
- Follow donor culture results for acute infections
- Donor history/serology
- Other transplant recipients with similar infections
- Types of donor derived infections
  - HSV
  - LCMV
  - WNV
  - HIV
  - Rabies
  - Trypanosoma cruzi



# Immunosuppression

- We make our patients susceptible to infections to prevent rejection
- Corticosteroids:
  - Broad inhibition of immune response (innate inflammatory response, phagocytic function, cellular immunity, possible antibody formation)
  - Hyperglycemia
  - Avoidance may reduce post-txp CMV infections (liver)
- Cytotoxic agents
  - Suppress bone marrow, peripheral blood counts
  - Cyclosporine, Tacrolimus, Mycophenolate mofetil, sirolimus

# Immunosuppression

- Patients have continued immunosuppression after initial prophylaxis
- Treatment of episodes of acute rejection, especially with high dose steroids “resets” the clock
- Increased risk of opportunistic infections after steroid boluses
- Toxicity associated with immunosuppressive therapy
  - Tacrolimus – renal toxicity, neurologic, diarrhea, diabetes
  - Mycophenolate – bone marrow suppression, diarrhea

# Immunosuppression

**TABLE 311-2** Biologic Preparations Used to Prevent or Treat Rejection

| AGENT  | ADVERSE EFFECTS  |
|--|--|
| <b>Polyclonal Antibodies</b>   |  |
| Antithymocyte globulins*   | Serum sickness, thrombocytopenia, lymphopenia (can last up to 2-3 yr with Thymoglobulin), increased risk of CMV, PTLD  |
| Anti-human thymocyte immune globulin (rabbit) (Thymoglobulin)            |  |
| Lymphocyte immune globulin, antithymocyte (equine) (Atgam)               |  |
| <b>Monoclonal Antibodies</b>   |  |
| Anti-CD25 (interleukin-2 receptor) antibodies†<br>Basiliximab (Simulect) | Hypersensitivity reactions, infection risk not significantly increased   |
| Anti-CD20 antibody‡<br>Rituximab (Rituxan)                               | Infusion reactions, hepatitis B virus reactivation   |
| Anti-CD52 antibody§<br>Alemtuzumab (Campath)                             | Infusion reactions, increased risk of CMV, <i>Pneumocystis jirovecii</i> pneumonia, invasive fungal infections, immunosuppression effects that can last up to 12 mo. |
| <b>Other Agents</b>  |  |
| Anti-B7 fusion protein (co-stimulation ligand)¶<br>Belatacept (Nulojix)  | Increased rate of Epstein-Barr virus-associated PTLD   |

CMV, cytomegalovirus; PTLD, post-transplantation lymphoproliferative disease.

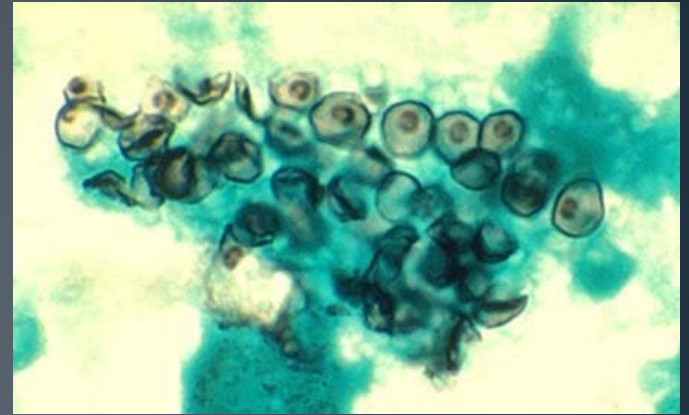
Mandell: Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 8<sup>th</sup> ed.; Chapter 311: Risk Factors and Approaches to Infections in Transplant Recipients. 3414-3424.

# Case #6

- A 39-year-old male is evaluated for acute rejection (Banff grade IA) of a transplanted kidney. PMH is significant for hypertension and autosomal-dominant polycystic kidney disease requiring transplantation 17 months ago. Donor and patient were seronegative for cytomegalovirus, and patient received standard prophylaxis (TMP-SMX and Acyclovir for 6 months) Post transplantation course has been complicated by previous episodes of rejection, last occurring 6 months ago. Current medications include amlodipine, tacrolimus, prednisone, and mycophenolate. He is scheduled to have his level of immunosuppression increased significantly (pulse IV methylprednisolone 3mg/kg daily x5 days, followed by taper to maintenance dose, and adjustment of maintenance immunosuppression as needed) for the current episode of rejection.
- Physical examination findings is relatively unremarkable. Laboratory studies show a leukocyte count of 5200/ $\mu$ L with 85% polymorphonuclear cells and 6% lymphocytes and a serum creatinine level of 2.0 mg/dL.
- For which infection should this patient receive prophylaxis at this time?
  - A) Aspergillus
  - B) Cytomegalovirus
  - C) Pneumocystis jirovecii
  - D) Mycobacterium avium complex

# *Pneumocystis jirovecii*

- Opportunistic fungal pathogen
- Primarily causes pulmonary disease
- In non-HIV patients who are immune suppressed, rapid development of disease
- Diagnose by immunofluorescent or silver stain
- Bactrim is treatment of choice
- Much lower incidence in post SOT due to Bactrim prophylaxis
- At higher risk when steroids are increased



Credits

Arthur Ammann, MD, Global Strategies for HIV Prevention

Description

Lung biopsy using silver stain to demonstrate organisms in tissue.

# Polyomaviruses – JC, BK

- Up to 90% adults seropositive for BK, 86% for JC
- Do not cause disease in immunocompetent persons
- BK – hemorrhagic cystitis in renal txp, HSCT
  - Up to 10% post renal txp have BK nephropathy
  - PCR for blood, urine. Cytopathic changes on biopsy.
  - Treatment – reduce immunosuppression
- JC – Lytic infection of oligodendrocytes in brain – leads to PML
  - PCR of CSF to diagnose
  - MRI with hyper intense white matter lesions on T2, flair. Hypo intense on T1
  - Lower immune suppression

## Case 7



A 45-year-old woman born in Thailand, with history of renal failure 2 months post renal transplant presents to the ED with complaint of fevers, cough, abdominal pain, and malaise for 1 week. She moved to the U.S. 30 years ago and denies any recent travel. She has been enjoying gardening now that she is no longer on dialysis. She is compliant with her immunosuppressive medications since her transplant. On physical examination, she appears ill and has diffuse crackles throughout bilateral lung fields, diffuse abdominal tenderness, and a maculopapular rash on her buttocks.

# Initial workup

Labs?

Other tests?

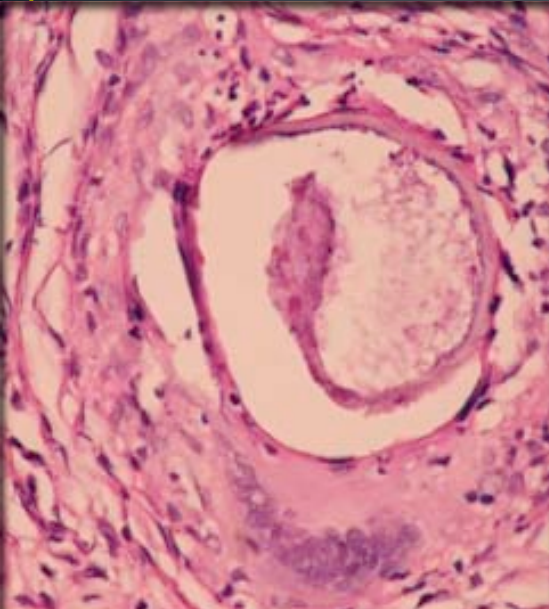
Key points in the history?

What is your differential diagnosis?

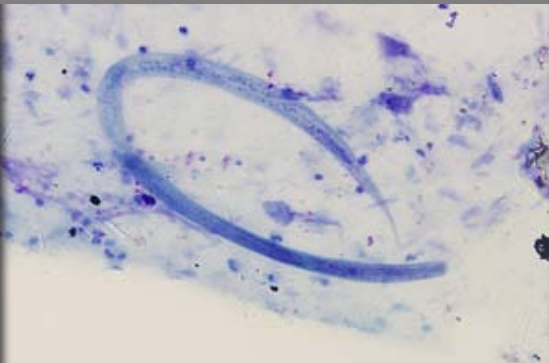
- WBC 8,000, Hg 12.9, PLT 450,000
- Peripheral smear:
- Eosinophils 8 %,
- Lymphocytes 10 %
- Monocytes 6 %
- Neutrophils 74 %
- ABG: pH 7.39, pCO<sub>2</sub> 35.2 mmHg, pO<sub>2</sub> 59 mmHg
- ESR 75 mg/dl
- HIV negative



# CASE 7: Workup



*Dogan et al. Pulmonary infection. Respiratory Medicine Case Reports 2014.*



*Image courtesy of CDC.gov*

Bronchoscopy with alveolar lavage is performed and cultures are negative for bacterial pathogens.

Microscopic examination and cytology is performed on the specimen and reveals the images on the left:

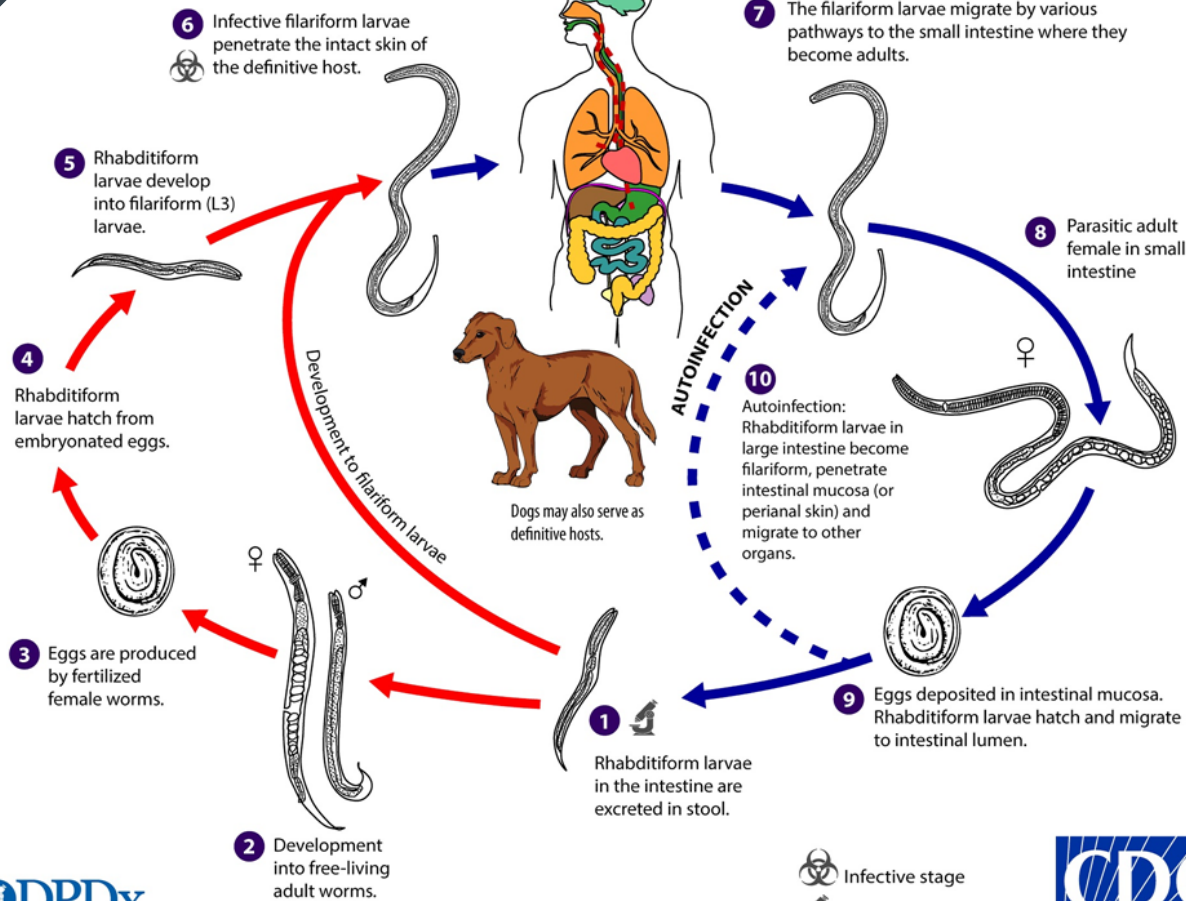
# Case 7: Question

What is the most likely diagnosis?

- A. Trichuris
- B. Ascaris
- C. Hookworm
- D. Strongyloides
- E. Trichinella

### Free-Living Cycle

### Parasitic Cycle



## Case 7 Strongyloidiasis

# Case 7: Roundworm infections

| Roundworm/<br>Nematode | Epidemiology                                       | Human Disease Manifestation                                       |
|------------------------|--|---|
| Enterobius             | Ingestion - human feces. Daycare.                  | Perianal pruritus   |
| Trichuris              | Ingestion - soil w/ human feces                    | Tenesmus  |
| Ascaris                | Ingestion - soil w/ human feces. Tropic/Subtropic. | Adult - intestinal obstruction; Larvae - transpulmonary migration |
| Hookworm               | Skin - soil w/human feces. Tropic/Subtropic.       | Adult - anemia; Larvae - transpulmonary migration                 |
| Strongyloides          | Skin - soil w/human feces. Latin America, SE Asia. | Larvae - transpulmonary migration, rash, hyperinfection syndrome  |
| Trichinella            | Eating undercooked pork.                           | Larvae - myalgia  |

# CASE 7 - *Strongyloides stercoralis*

## **Epidemiology:**

- Found in the **soil** in rural subtropical and temperate regions
- Transmission may be through ground-skin, fecal-oral, contact with infected fomites
- Infections may be chronic

## **Clinical Manifestations:**

- Rash/pruritus at the site of inoculation Dry cough, wheezing
  - (may worsen w/ steroids)
- “Larva currens”
- GI symptoms – diarrhea/constipation/pain
- Rash with autoinfection....

## **Diagnosis:**

- Stool O&P
- Serology

# CASE 7 - *Strongyloides stercoralis* –

Larva Currens

Edematous rash pathognomonic for strongyloidiasis

Represent migrating larva in the skin, may develop at rate of 5-15cm/hr



Corte, Liliam Dalla et al. "Simultaneous Larva Migrans and Larva Currens Caused by *Strongyloides stercoralis*: A Case Report." *Case Reports in Dermatological Medicine* 2013 (2013): n. pag.

# Fever workup in transplant recipient

- Comprehensive history and exam
- CBC w/diff, CMP, CXR, UA, Blood and urine cultures
- Symptom/timeline appropriate workup
  - ?Respiratory pcr
  - CMV pcr
  - CT scanning or other imaging
- LP if headache, neurologic deficits
- Fevers >7days?
  - CMV, EBV, HHV6
  - Fungal?
  - Mycobacterial or other atypical
  - PJP, cryptococcal disease, tick borne illness
  - TB

# Fever workup in transplant recipient

- Consider non infectious etiology of fever
  - Rejection
  - Drug reaction
- Some infections may present without fevers
  - Are they receiving steroids?
  - PJP – cough, shortness of breath
  - Cryptococcal infection – headache, non responsive cellulitis
  - PML – neurologic deficits



# Post-transplant Prophylaxis

- Usually protocol, risk factor dependent
- TMP/SMX well tolerated, generally at least 3 months
  - Protection against PJP, Toxoplasma, Nocardia, Listeria, Legionella
  - Alternatives – Dapsone, Atovaquone, inhaled pentamidine
- Valganciclovir
- Fluconazole
- Fluoroquinolone (BMT)

# Vaccination

- Evaluate before transplant, less response once immunosuppressed
  - MMR
  - Tdap
  - HAV/HBV
  - Pneumonia (Pneumovax, pneumovax)
  - Influenza
  - ?Varicella (not after transplant)

# Questions?



# References

- 1. Mandell: Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 8<sup>th</sup> ed.; Chapter 311: Risk Factors and Approaches to Infections in Transplant Recipients. 3414-3424.
- 2. <https://www.unos.org/data/>
- 3. <https://optn.transplant.hrsa.gov/>
- 4. *Bone Marrow Transplantation* (2008) **42**, S3–S5; doi:10.1038/bmt.2008.102
- 5. [http://bloodcell.transplant.hrsa.gov/research/transplant\\_data/transplant\\_activity\\_report/index.html](http://bloodcell.transplant.hrsa.gov/research/transplant_data/transplant_activity_report/index.html)
- 6. Fishman, J. *Infection in Solid-Organ Transplant Recipients*. *N Engl J Med* 2007;357:2601-14.
- 7. Nasa M, Sharma Z, Sud R, Lipi L. Cytomegalovirus infection of gastrointestinal tract . *Community Acquir Infect* [serial online] 2016 [cited 2016 Aug 7];3:4-9. Available from: <http://www.caijournal.com/text.asp?2016/3/1/4/179226>